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Introduction to Transgenic Plant

Zoology Assignment → Shivaji College, (DU)

Topic:- Production and Application of Transgenic Plant

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Submitted by: GunGun Gang

I-Sem - 'A'

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INTRODUCTION OF TRANSGENIC PLANTS.

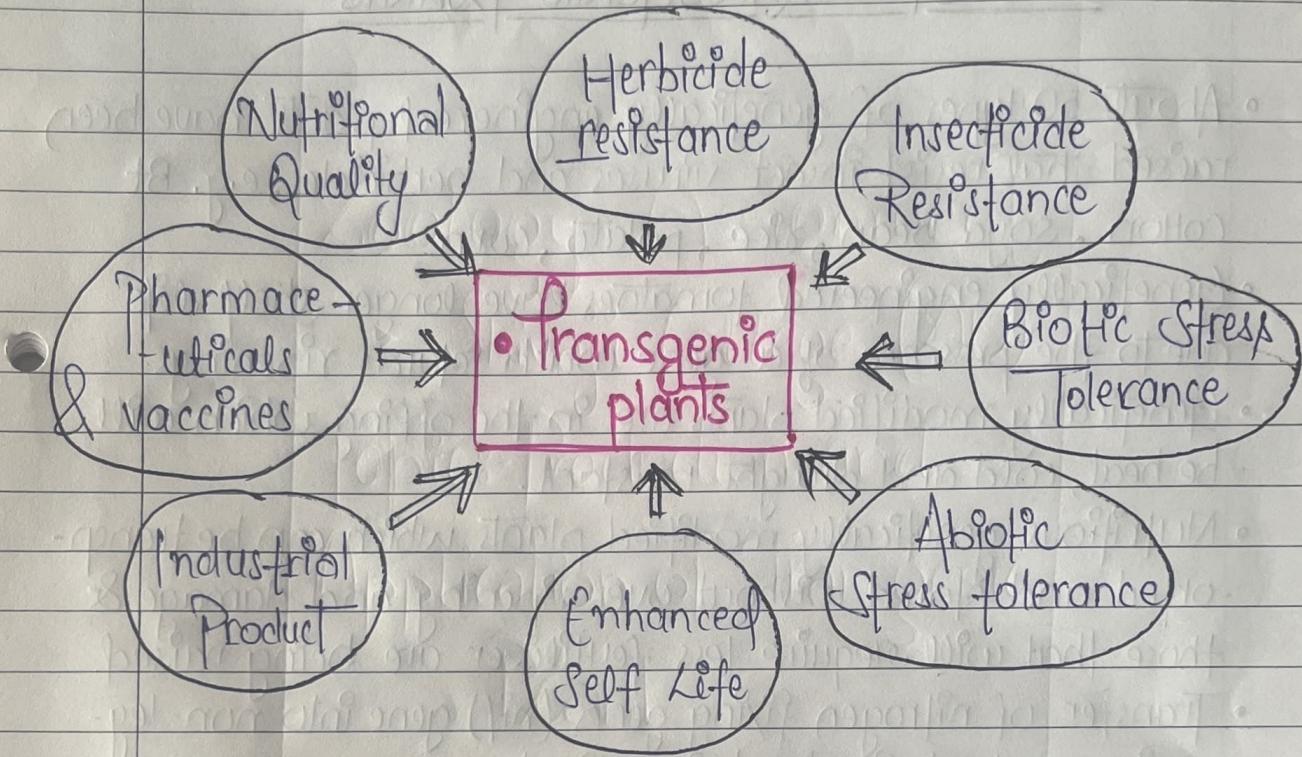
- Transgenic plants are plants in which one or more genes from another species have been introduced into the genome using genetic engineering processes.
- The aim is to introduce a new trait to the plant which doesn't occur naturally in the species.
- A transgenic plant consists a gene or genes that have been artificially inserted. The inserted gene sequence is known as transgene, it may come from an unrelated plant or from a completely different species.
- Techniques involve the biolist method - in which a heavy metal is coated with plasmid DNA is shot into cell - and Agrobacterium tumefaciens mediated transformation.
- The word 'Transgenic' stands for any external genetic features artificially introduced into the genome of another organism to get desired features.
- The 'Flavr Savr' tomato was the first transgenic plant that made its appearance in the market, developed by Celgene.
- With the advent of new molecular biology techniques such as crispr-cas 9, the future of transgenic plants has unlimited potential.
- 1st transgenic plant produced which is an antibiotic resistance tobacco plant.
- 1st successful plant genetic engineering experiment using the caulimovirus vector.
- 1st pesticide producing crop, Bt potato was approved by U.S. Environmental Protection Agency.
- GloGen rice - with β -carotene developed with increased nutrient value.
- 1st genetically engineered crop developed by Robert Fraley,

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Marc Van Montagu and Mary Dell Chilton was awarded World Food Prize.

Why TRANSGENIC PLANTS ?



Production of TRANSGENIC PLANTS

- So to create a transgenic plant, first, we need to identify a helpful gene (this acts as our gene of interest) in an organism we want to insert into our plant.
- This gene is then extracted with the help of restriction endonuclease enzymes (molecular scissors) and inserted into the target genomes.
- These insertions can be carried out by several methods such as:
 - Particle Gun or gene gun or biotics,
 - PEG (polyethylene glycol mediated transformation).
 - electroporation.
 - Agrobacterium-mediated gene transfer.

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Generally, a small section of cells take up the gene of interest, and thus they are selected and regenerated into transgenic plants via tissue culture techniques.

• USES OF TRANSGENIC PLANTS

- About 50 types of genetically engineered plants have been raised that resist insect viruses and herbicides, e.g. Bt cotton, soya bean, rice, potato, corn.
- Genetically engineered tomatoes have longer shelf life due to the presence of a gene which retards ripening.
- Genetically modified plants are in the offing which will be heat, cold and drought resistant.
- Nutritious genetically modified plants which can be transported and stored without over ripening and damage & those that will require less fertilizer are being raised.
- Transfer of nitrogen fixing genes (*nif*) gene into non-leguminous plants is also being tried.
- Plants are also being genetically engineered to produce in their seeds proteins (e.g. Hormones) which are of interest to humans
- A weed name - mouse-eared cress has genetically engineered to produce a biodegradable plastic poly hydroxyl butyrate or PHB inside its cell granules.

• ADVANTAGES OF TRANSGENIC PLANTS.

for the producers of the new varieties.

- A high efficiency in plants is obtained.

for farmers

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- Process of press destroying is simplified, therefore less damage.
- Production output is increased as well as profits of the transgenic culture, even the obtaining cost of the GMO is rather high.

for industry

- As in the case of modified starch, low lignin content wood (in this case paper manufacturing is less pollutant), human protein production (for therapeutic aim).

for consumers

- More nutritious
- Fruits and vegetables are delayed maturation can easily be stored, with minimum losses.
- In future, transgenic plants contain high content of vitamins, minerals, essential amino acids by using the vaccine products of plant, the rice enriched in pro-vitamin A, etc.

for the environment and human future

- Imply lower pollution
- Higher agriculture productions.

• DISADVANTAGES OF TRANSGENIC PLANTS

Damage to human health

- Allergies
- horizontal transfer and antibiotic resistance.
- eating foreign DNA
- changed nutrient values.
- Large genomes

(polyploid → presence of many genomes in one cell)

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Damage to the natural environment

- Crop to weed gene flow
- leakage of GM protein into soil
- reductions in pesticides spraying : are they real ?

Disruptions of current practices of farming and food

production in developed countries.

- Crop-to-crop gene flow

Disruptions of traditional practices and economics in less developed countries.

- Lack of research on consequence of transgenic crops.

PLANT GENE TRANSFER METHODS

**INDIRECT
METHODS
(vector-based)**

**DIRECT
METHOD
(vector-less)**

**IN-PLANTA
TRANSFORMATION**

✓ • Agrobacterium mediated transformation.

• Physical methods
→ Particle bombardment
→ Electroporation
→ Microinjection
→ Liposome mediated

• floral Dip
• Vacuum infiltration
• Agro infection

• DNA transfer
→ Silicon Carbide fibre mediated DNA transfer
• Chemical method
→ PEG-mediated DNA transfer.

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AGROBACTERIUM - MEDIATED TRANSFORMATION

→ Below are the basic steps in the Agrobacterium-mediated Plant Transformation Process:

1. Isolate genes of interest from the source organism.
2. Create a functional transgenic construct that include:
 - (a) The gene of interest
 - (b) Expression Promoters
 - (c) Codon optimization that increase protein production
 - (d) Marker genes to track gene expression in the host plant
3. Insert transgene into Ti plasmid.
4. Insert T-DNA containing plasmid into Agrobacterium.
5. Mix newly transformed agro cells with plant cells to allow transfer of the T-DNA into the plant chromosomes.
6. Regenerate the transformed cells into genetically modified plants.
7. Test at various stages to ensure trait performance.

- Agrobacterium tumefaciens and Agrobacterium rhizogenes are soil-borne, Gram-negative bacteria. These are phytopathogen (that cause infection in plants) and as they are treated as the nature's most effective plant genetic engineer. A. tumefaciens induces crown gall disease and A. rhizogenes that induces hairy root disease in plant.

Crown Gall Disease-Ti plasmid.

- A. tumefaciens infects wounded or damaged plant tissues, it induces the formation of a plant tumor called crown gall.
- The entry of bacterium into the plant tissues is facilitated by the release of certain phenolic compound by wounded sites.
- It is a rod-shaped, Gram-negative soil bacterium, the causal agent of crown gall disease in over 140 species of dicots.

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formation of a crown Gall Tumor.

- Crown gall formation occurs when the bacterium releases its Ti plasmid into the plant cell cytoplasm. A fragment of Ti plasmid, referred to as T-DNA, is actually transferred from the bacterium into the host where it's integrated into the plant cell chromosomes. Thus, crown gall disease is a naturally evolved genetic engineering process.
- The T-DNA carries gene that code for proteins involved in the biosynthesis of growth hormones (auxin and cytokinin) and novel plant metabolites namely opines- amino acid derivatives and agropines- sugar derivatives.
- The growth hormones cause plant cells to proliferate and form the gall while opines and agropines are utilized by *A. tumefaciens* as source of carbon and energy. Thus, *A. tumefaciens* genetically transforms plant cell and creates a biosynthetic machinery to produce nutrients for its own use.
- As the bacteria multiple, & continue infections, crown gall develops which is a visible mass of the accumulated bacteria and plant material. Crown gall formation is the sequence of the transfer, integration & expression of gene of T-DNA (or Ti plasmid) of *A. tumefaciens* in the infected plant.

Organization of Ti plasmid:

- The Ti plasmids (approx 900 kb each) exist independent replicating circular DNA molecules with the Agrobacterium cell. The T-DNA is variable in length in range of 12 to 24 kb which depend on the bacterial strain from which Ti plasmids come.

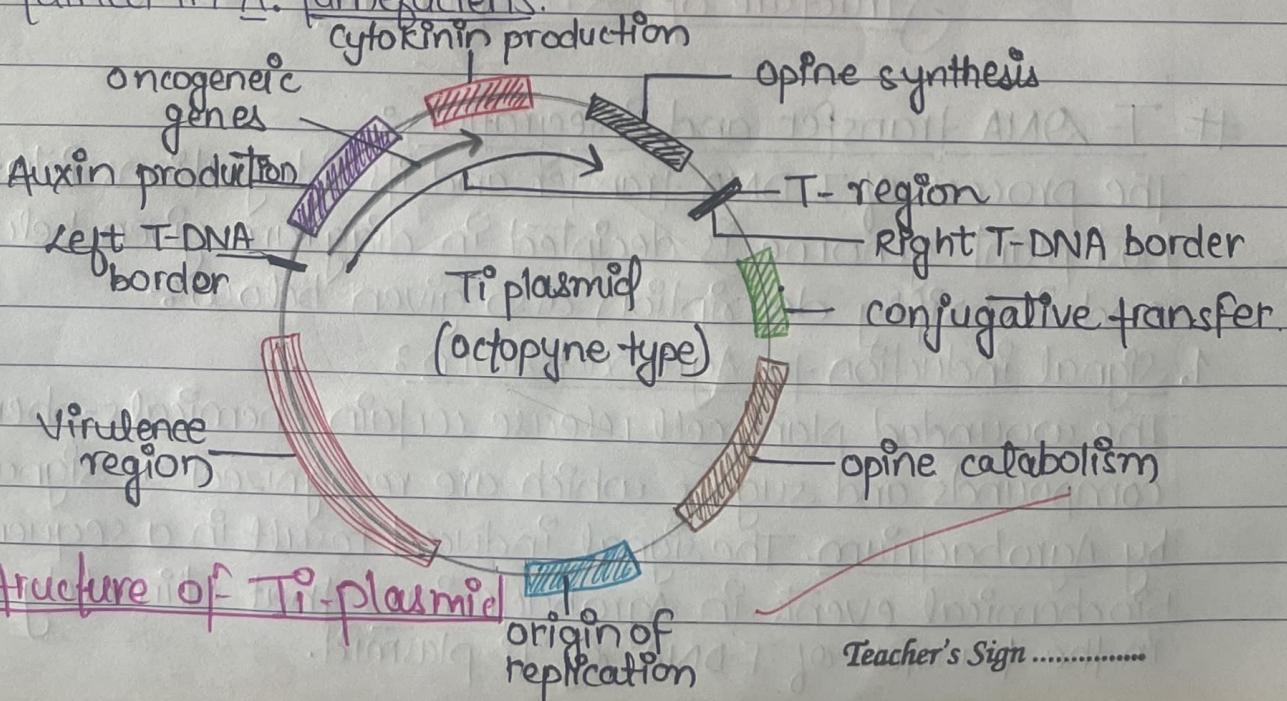
→ Nopaline strains of Ti plasmid have one T-DNA with length of 90 kb while octopine strains have two T-DNA regions referred to as TL and TR that are respectively 14 kb & 7 kb in length.

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The Ti plasmid has three important regions :-

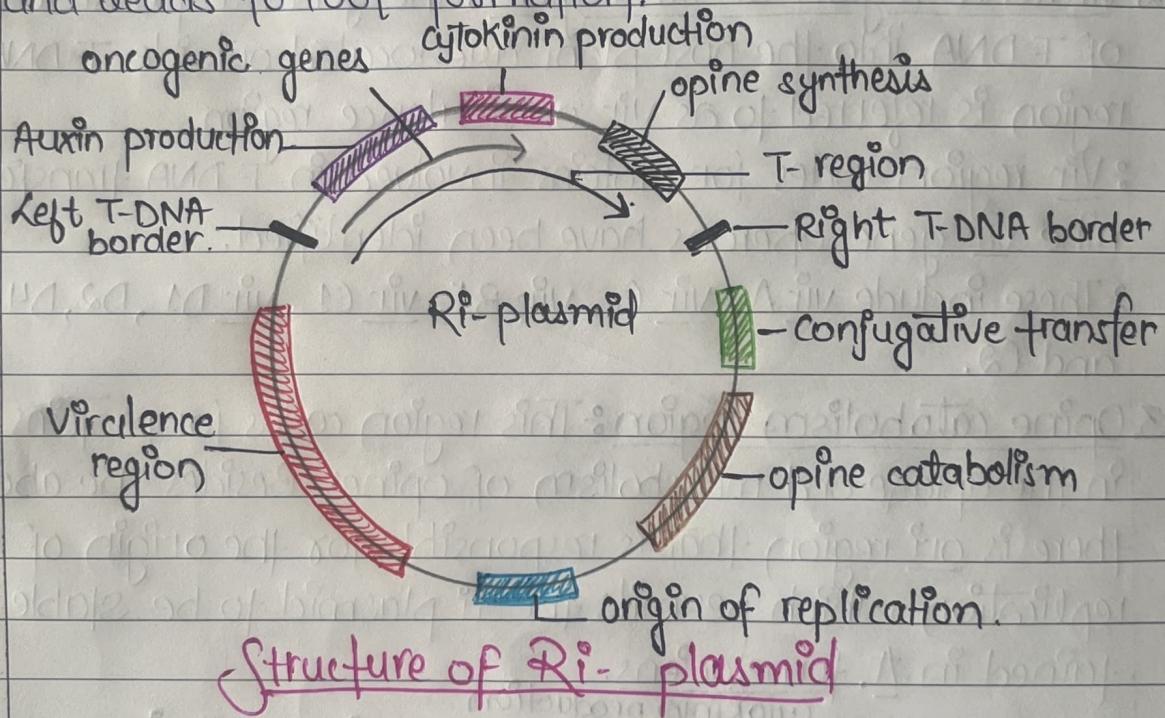
1. T-DNA regions : This region has the genes for the biosynthesis of auxin (aux), cytokinin (cyt) and opine (ocs) and is flanked by left and (cytokinin) right borders.
 - These three genes - aux, cyt and ocs are referred to as oncogenes, as they are the determinants of the tumor phenotype.
 - T-DNA borders - A set of ~4kb sequences present on either side (right and left) of T-DNA are also transferred to plant cell.
 - It is now clearly established that the right border is more critical for T-DNA transfer and tumorigenesis.
2. Virulence region or vir region : The genes responsible for transfer of T-DNA into the host plant are located outside T-DNA and the region is referred to as vir or virulence region.
 - Vir region codes for proteins involved in T-DNA transfer. At least nine vir-gene operons have been identified.
 - These include virA, virG, virB1, virC1, virD1, D2, D4 and virE1 and E2.
3. Opine catabolism region : This region codes for proteins involved in the uptake and metabolism of opiates. Besides the above three, there is ori region that is responsible for the origin of DNA replication which permits the Ti plasmid to be stable maintained in *A. tumefaciens*.



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Hairy Root Disease of *A. Rhizogenes* - R^i plasmids:

- *Agrobacterium rhizogenes* can infect plants that cause hairy root disease. The plasmids of *A. rhizogenes* are referred to as R^i plasmid (Root-Inducing plasmids). These are of different types. Some of the R^i plasmid strain possess gene that are homologous to T^i plasmid eg. auxin biosynthesis genes.
- Instead of virulence genes, R^i plasmid gene contain a series of open reading frames on the T-DNA.
- The product of these genes are involved in the metabolic of plant growth regulations which gets sensitized to auxin and leads to root formation.



T-DNA transfer and integration:

The process of T-DNA transfer and its integrations into the host plant genome is depicted in diagram given below and is briefly description also given below:-

1. Signal induction to *Agrobacterium*:

The wounded plant cell release certain chemicals-phenolic compounds and sugars which are recognized as signals by *Agrobacterium*. The signal induced result in a sequence of biochemical events in *Agrobacterium* that ultimately help in the transfer of T-DNA of T^i -plasmid. Teacher's Sign

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2. Attachment of Agrobacterium to plant cell:

- They attach to the plant cell through polysaccharides, particularly cellulose fibres produced by the bacterium.

3. Production of Virulence proteins:

- As the signal induction occurs in the Agrobacterium cells attach to plant cell, a series of events that result in the production of virulence protein. To start with, signal induction by phenolics stimulates vir A which in turn activates (by phosphorylation) vir C. This induces expression of virulence gene of Ti-plasmid to produce the corresponding virulence protein (D1, D2, D3, E2, B). Certain sugars (eg. glucose, galactose, xylose) that induce virulence gene have been identified.

4. Production of T-DNA Strand:

- The right and left borders of T-DNA are recognized by vir D1 / vir D2 proteins. These proteins are involved in the production of single-strand T-DNA, its protection and export to plant cells. The ss T-DNA gets attached to vir D2.

5. Transfer of T-DNA out of Agrobacterium:

- The ss T-DNA-vir-D2 complex in association with vir G is exported from the bacterial cell. Vir B product from the transport apparatus.

6. Transfer of T-DNA into plant cell and Integration:

- The T-DNA-vir-D2 complex cross the plant plasma membrane. In the plant cell, T-DNA gets covered with vir E2. This covering protects the T-DNA from degradation by nucleases. Vir D2 and vir E2 interact with a variety of plant proteins which influence T-DNA transport and integration.
- The T-DNA-vir D2-E2 plant protein complex enter the nucleus through nuclear complex pore. Within the nucleus, the T-DNA gets integrated into the plant chromosome.

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through a process referred to illegitimate recombination.

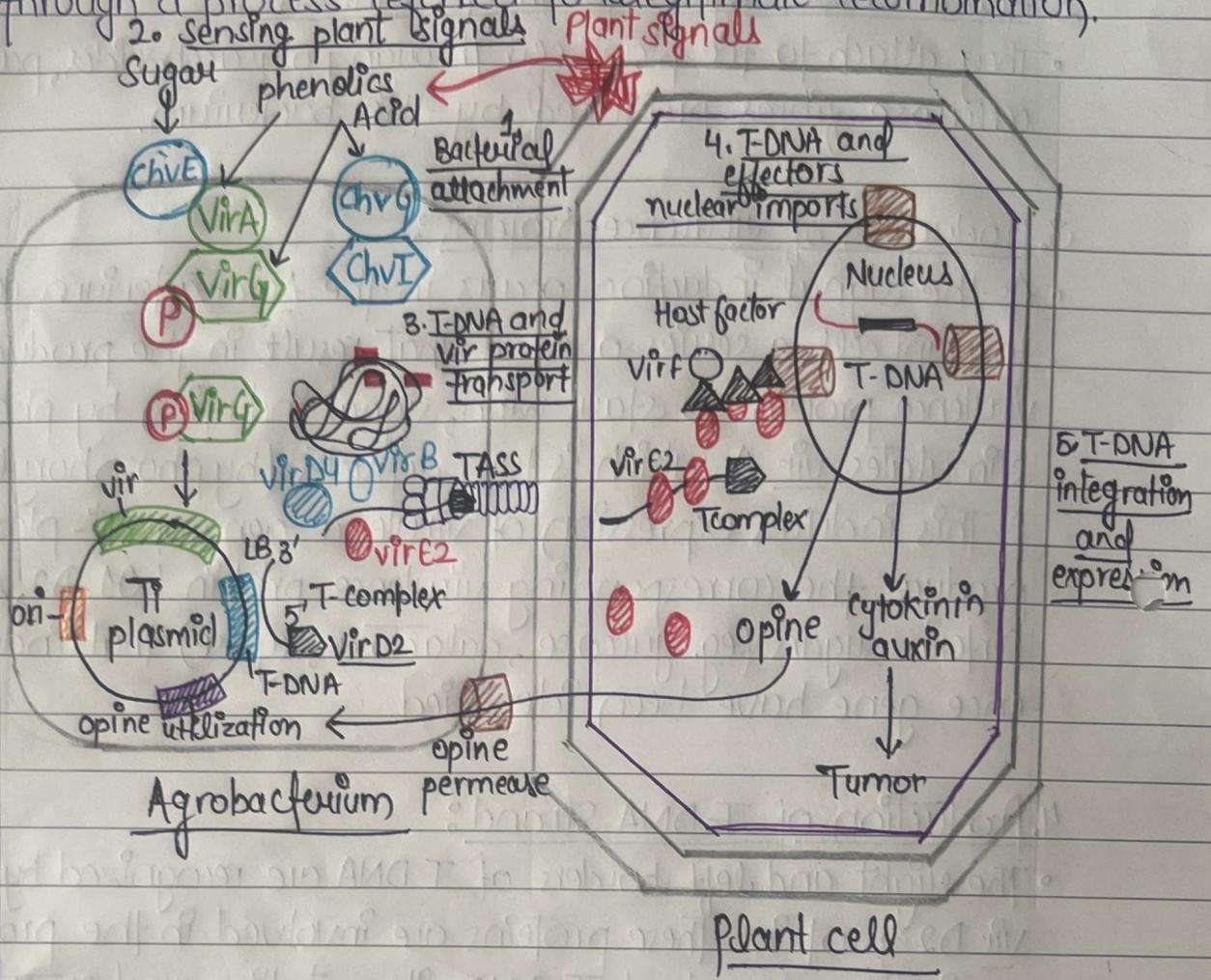


fig:- Process of T-DNA transfer and its integration into host plant genome

Modification of Ti plasmid:

- Two types of Ti-plasmid-derived vectors are used for genetic transformation of plants.
- Co-integrate vector
 - Binary vectors
 - In the system, the disarmed and the modified Ti-plasmid combines with an intermediate cloning vector to produce a recombinant Ti-plasmid.

Production of disarmed Ti plasmid:

In these Ti plasmid, the oncogenes located in the T-DNA region have been replaced by exogenous DNA.

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- Examples of these vectors include :

1. SEV series :- The right border of the T-DNA together with the phytochrome gene coding for cytokinin and auxin are removed and replaced by a bacterial kanamycin resistance gene while the left border and a small part of the left segment (TL) of the original T-DNA (referred to as left inside homology (L IH)) are left intact.
2. pGV series : The phytochrome gene are excised and substitute by part of pBR322 vector sequence. The left and right border sequence as well as the nopaline synthase gene of the Ti plasmid are conserved.

Construction of Intermediate vectors

- These are small pBR322-based plasmids consist a T-DNA region. They are used to overcome the problem derived from the large size of disarmed Ti plasmids and their lack of unique restriction sites.
- Intermediate vector are replicated in E. coli and are transferred into Agrobacterium by conjugation.
- They cannot replicate in A. tumefaciens and therefore, carry DNA segment homologous to the disarmed T-DNA to permit recombination to form a co-integrated T-DNA structure.

Helper vectors

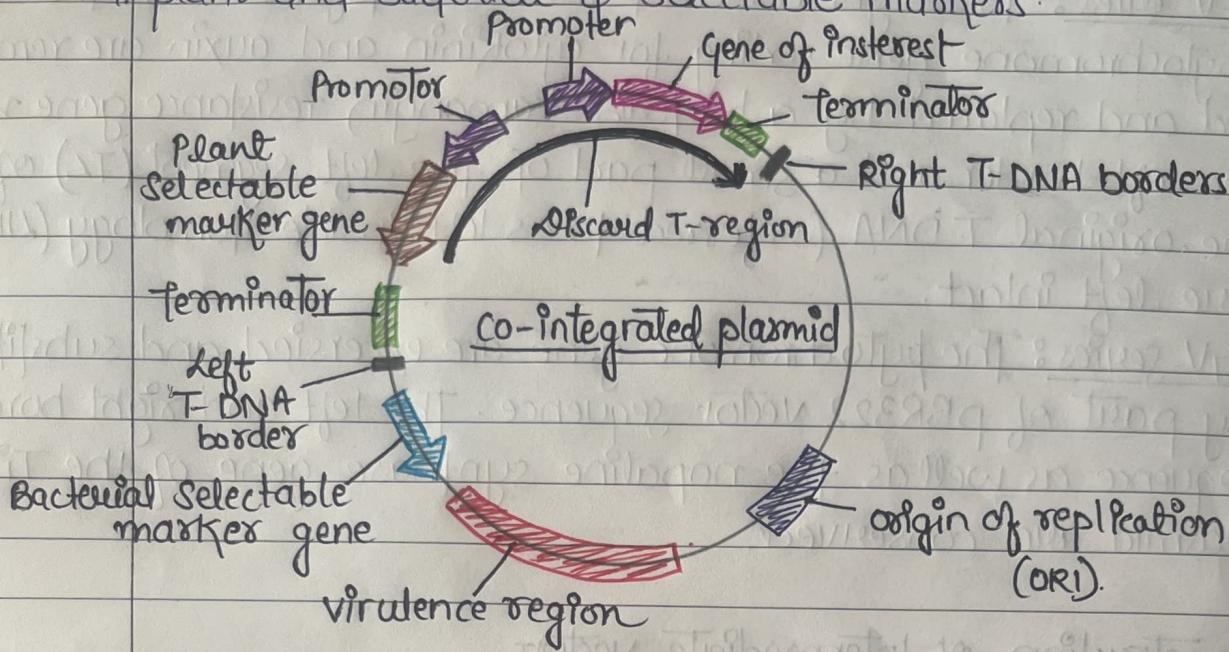
- These are small plasmids maintained in E. coli that contain transfer (tra) and mobilization (mob) genes, which allow the transfer of the conjugation-deficient intermediate vector into Agrobacterium.
- A resulting co-integrated plasmid assembled by in vitro manipulation normally contains :
 - the vir genes
 - the left and right border, T-DNA.

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3. an exogenous DNA sequence between the two T-DNA border.
4. plant and bacterial ♂ selectable markers.

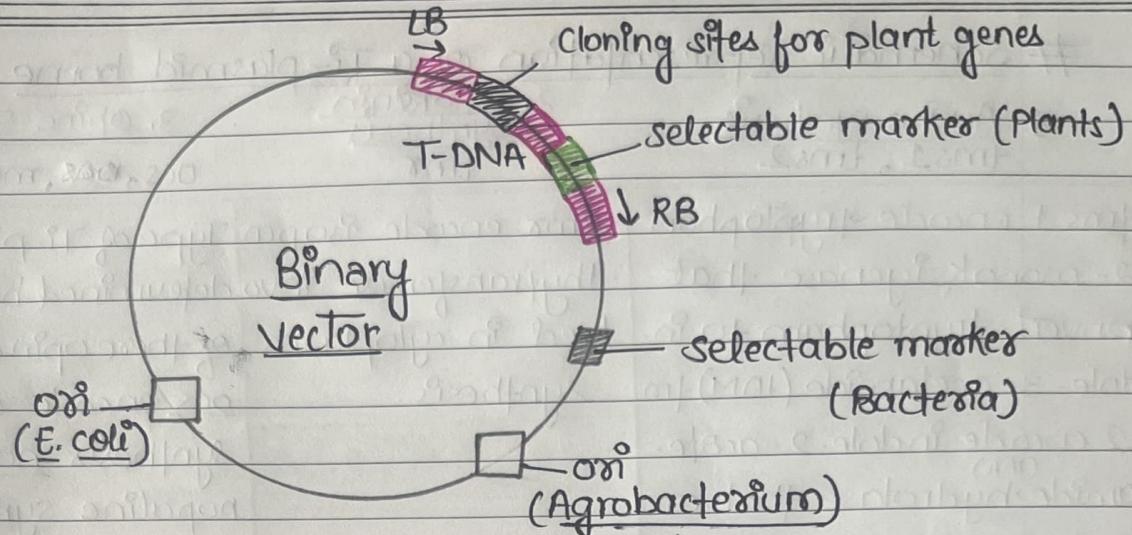


Binary Vector

- The binary vector system consist of an Agrobacterium strain along with a discard Ti plasmid called vir helper plasmid
- It may be noted that both of both them aren't physically linked. A binary vector with T-DNA can replicate in E.coli and Agrobacterium
- The binary vector has the following components:
 - Left and right border that delimit the T-DNA regions.
 - A plant transformation marker (PTM) e.g. npt II that confer Kanamycin resistance in plant transformed cells
 - A multiple cloning sites (MCS) for introducing target/foreign genes
 - A bacterial resistance marker eg. tetracycline resistance gene for selecting binary vector colonies in E.coli and Agrobacterium
 - oriT sequence for conjugal mobilization of the binary vector from E.coli to Agrobacterium.
 - A broad host range origin of replication such as RK2 that allow the replication of binary vector in Agrobacterium.

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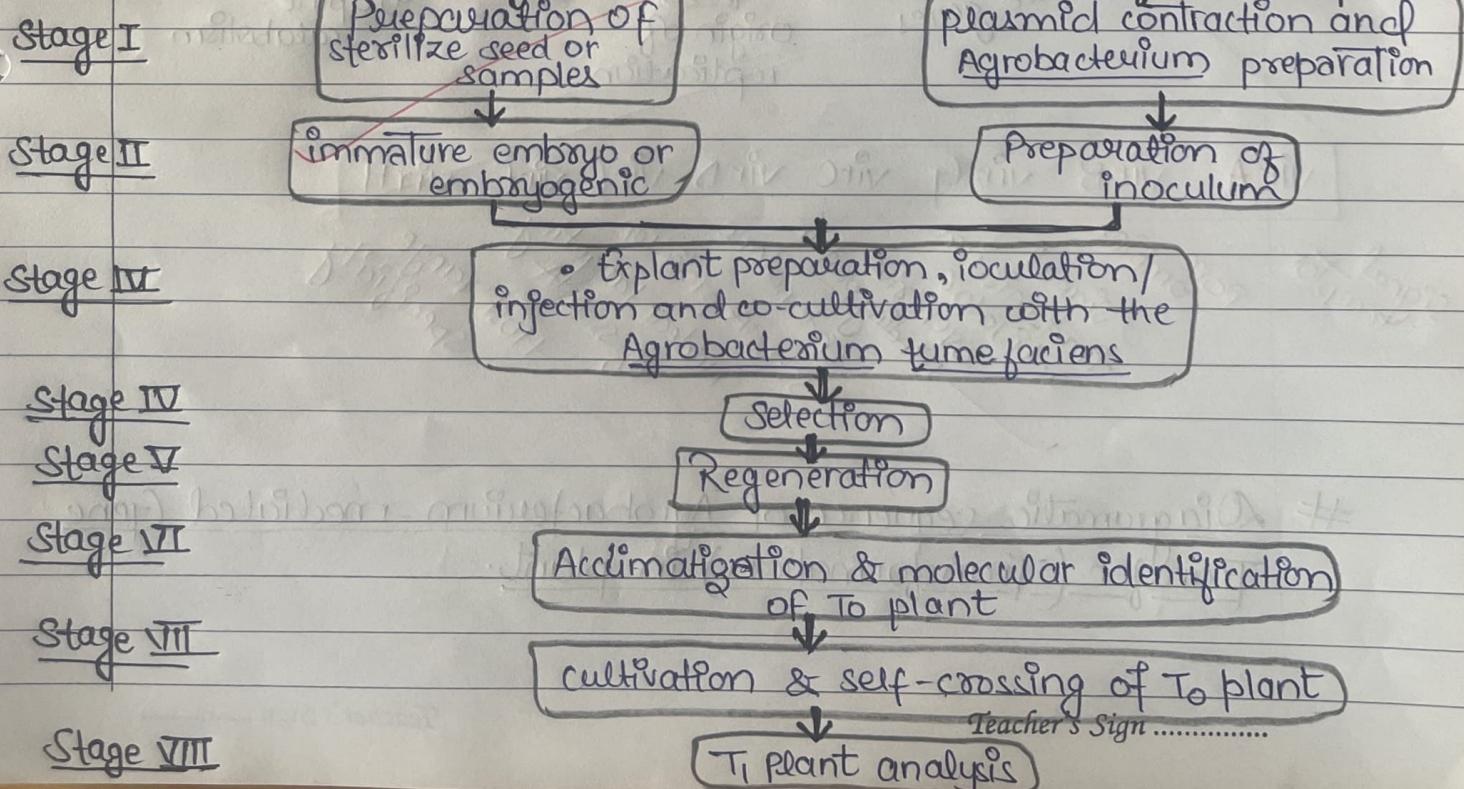
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Advantages of Agrobacterium mediated gene Transfer

1. Simple and comparatively less expensive
2. High transformation efficiency
3. Transgenic crops obtained have better fertility percentage
4. Protocols for both dicotyledons and monocotyledons are available
5. Relatively large length DNA segment can be transferred.

The protocol of Agrobacterium-mediated transformation.



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Schematic representation of a Ti-plasmid borne.

1 Auxine synthesis
tms1, tms2

2 Cytokinin synthesis
tmr

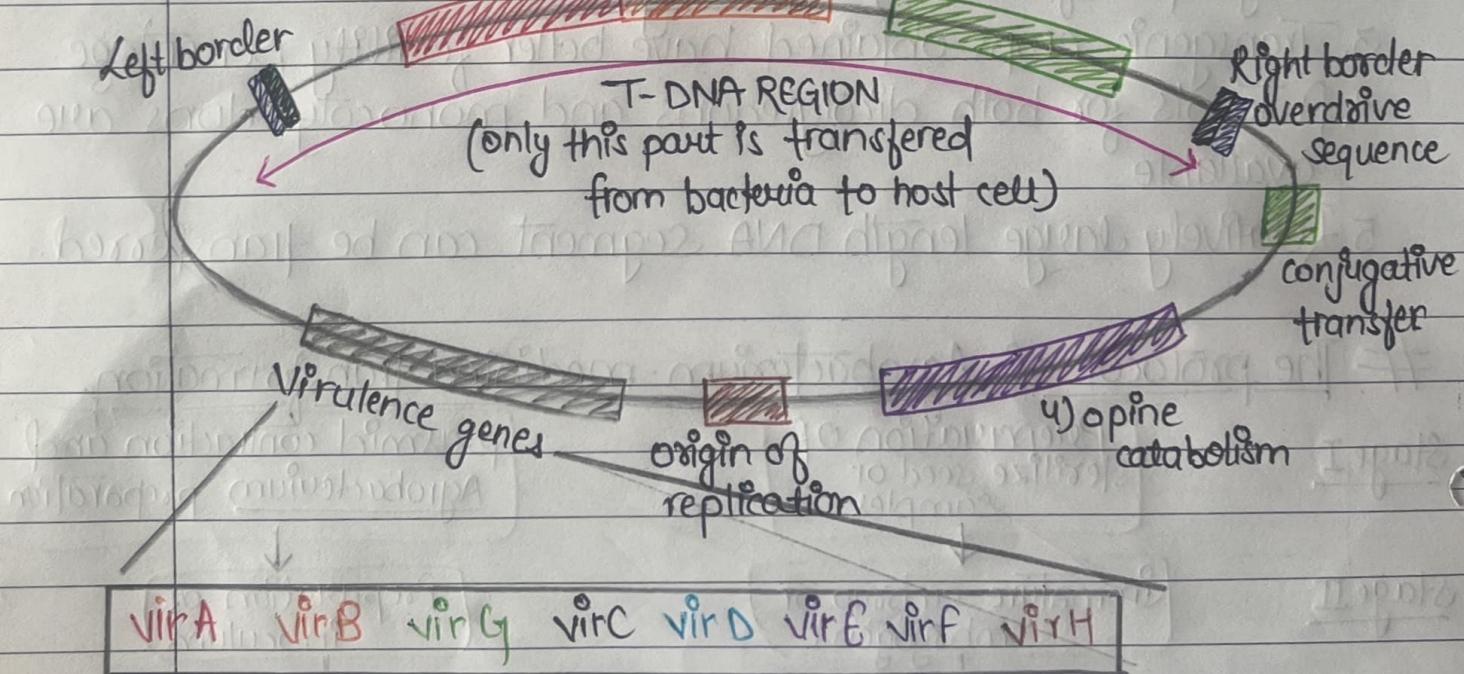
3. opine synthesis
ocs, nos, mas, ags.

tms1 encode tryptophan-2-monooxygenase that converts tryptophan to indole-3-acetamide (IAM). tms2 encode indole-2-acetamide hydrolase that converts IAM to IAA.

tmr encode 9-isopentenyltransferase involved in cytokinin synthesis.

Type of Ti-plasmid are determined by the type of these opine produced.
ocs encode octopine synthesis
nos encode nopaline synthase
mas encode mannopine synthase
ags encode agropine synthase.

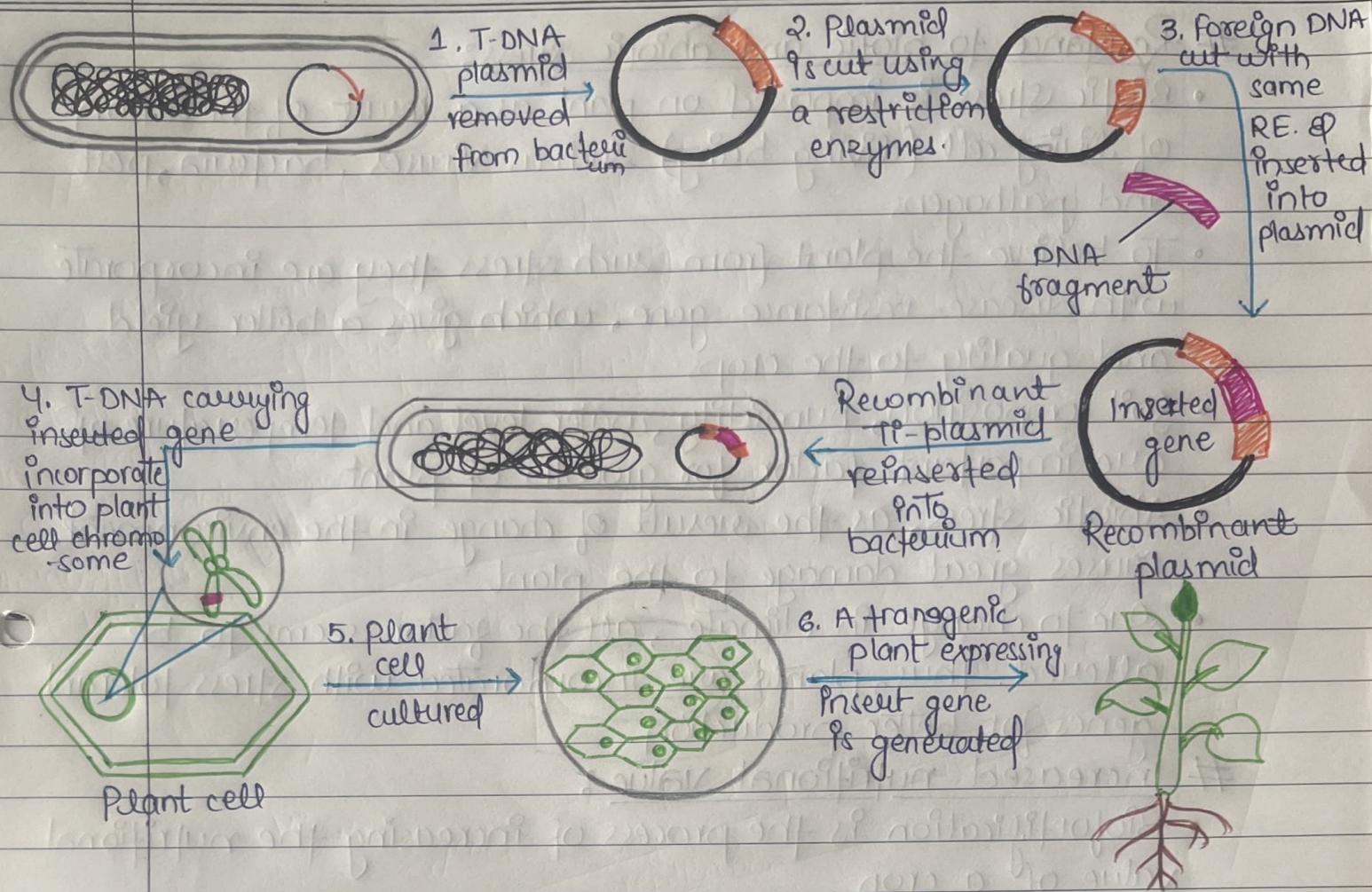
oncogenes (leads to tumor formation)



Diagrammatic sequence of Agrobacterium-mediated Gene Transfer (Transformation) in plant.

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APPLICATION OF TRANSGENIC PLANTS

Transgenic plants have various applications

Resistance to
Biotic Stress

Resistance to
Abiotic stress

Improvement of
crop yield & quality

Production of
low-cost
pharmaceuticals

1. Insect Resistance
2. Virus Resistance

3. Fungal and
Bacterial
Resistance

1. Herbicide
Resistance
2. Glyphosate
Resistance

1. Extended shelf
life of fruits
2. Improved
nutrition
3. Improved
coloration

1. Edible
vaccines
2. Essential
proteins.

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Resistance to biotic and abiotic stress:

- Biotic stress is imposed on plant as a result of the action of living cell (living being) such as virus, bacteria, pest and pathogens.
- To relieve the plant from such stress they are incorporate with disease-resistance gene, which gives a better yield and quality to the crops.
- Soil composition, humidity, water level and temperature of are important factor for growth of plant
- Abiotic stress, as the result of change in the environment, causes great damage to the plant.
- Due to change in climate, all the factor seems to be altered. Thus, plants are incorporated with stress-tolerant genes for better production.

Increased nutritional value:

- Biofortification is the process of increasing the nutritional value of a crop.
- Malnutrition is a common problem in developing countries.
- As a solution, plants are engineering to produce crops of better nutritional value.

factories of production of recombinant proteins

- Recombinant (DNA) human protein have been produced using animal and microorganism, system but due to some shortcoming it has been shifted to the plant system.
- Vaccines and antibiotics have been obtained from transgenic plant
- However, this application is still in the development stage and has not been commercialised yet.

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Insect Resistance Plants.

- It is estimated that about 15% of the world's crop crop yield is lost due to insect or pests.
- Bacillus thuringiensis was first discovered by Ishiwaki in 1901. It is a gram negative soil bacterium.
- Most of the Bt toxins are active against Lepidopteron larvae, while some of them are specific against Dipterans and the Coleopteran insects.
- Different cry proteins produced by Bacillus:
 - Cry I : kill butterflies and moth
 - Cry II : kill butterflies and flies.
 - Cry III : kill beetles.
 - Cry IV : kill only flies.
- Plant made only low levels of toxin because they are designed to express well in bacteria and not in plants as they are produced from bacterium.
- Insect toxin gene was altered by changing many bases of the third position of the redundant codon to improve its toxicity.

Bt toxin.

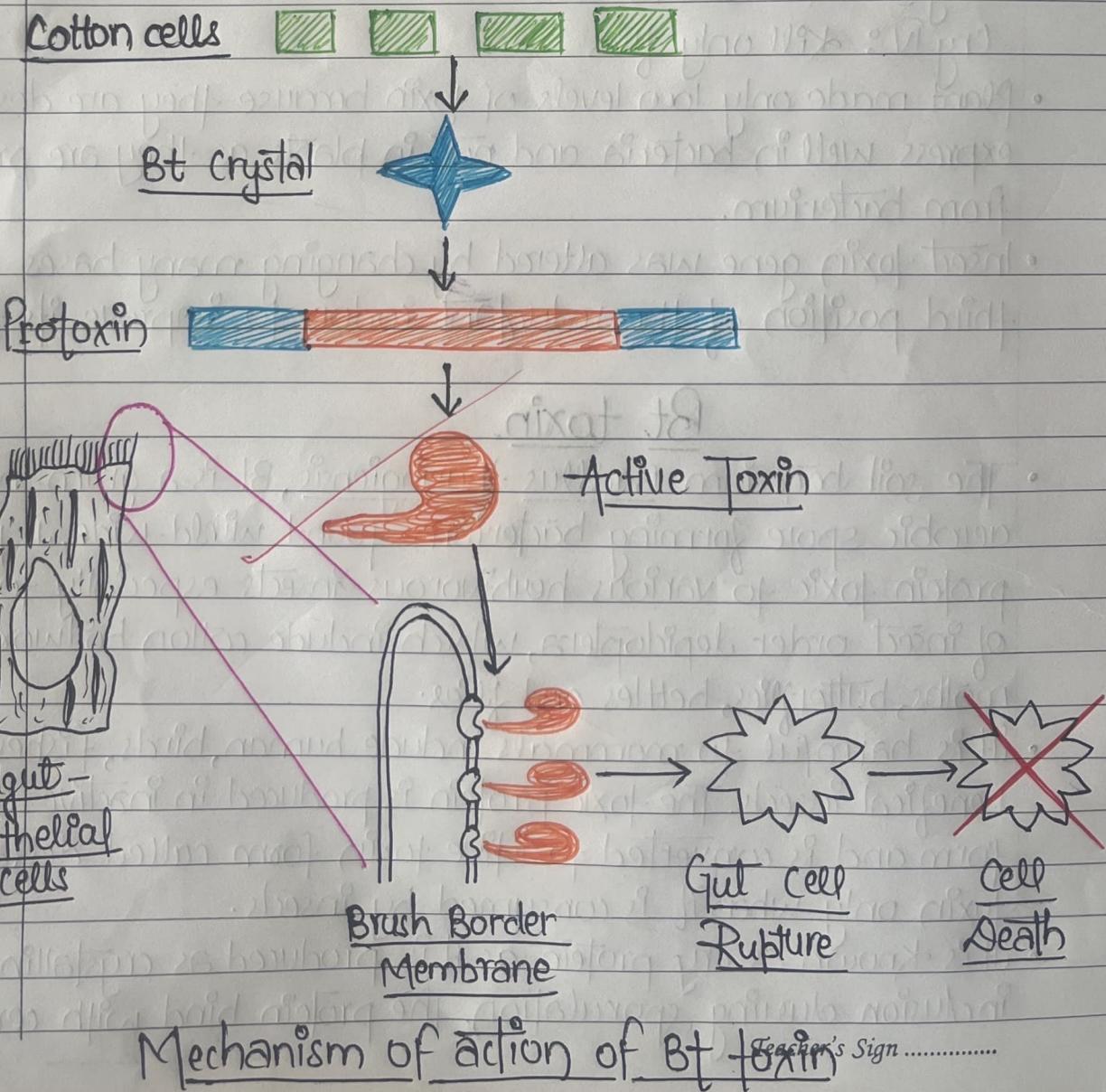
- The soil bacterium Bacillus thuringiensis, Bt is a gram-negative aerobic spore forming bacterium found worldwide. It produce protein toxic to various herbivorous insects especially to larvae of insect order Lepidoptera, which include cotton bollworms, moths, butterflies, beetles and flies.
- It is harmful to mammals include human, birds, fishes, or the beneficial insects. The toxic protein is produced in inactive crystalline form and is converted to active toxin form called delta endo-toxin only when it is consumed by insects.
- It is known as cry protein as it is produced as crystalline protein inclusion during sporulation. The protein bind with certain

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receptors (aminopeptidase N (APN) receptor & cadherin-like receptors) in insect's intestine and causes death of insects. Lysing of epithelial midgut cell.

- The toxin destroy gut of insect, ultimately rear leading to death of insect. The toxic protein is also known as Bt protein as it is produced by Bacterium Bacillus thuringiensis.
- In USA, it is registered as a biopesticide but the performance of Bt insecticide on cotton plant was limited. The insecticide can be degraded as light, heat, UV, high pH and desiccation.
- Even the area where the pest of cotton (boil worms) feeds are difficult to treat. The insect must eat sufficient treated plant to accumulate lethal dose of toxin.



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B. thuringiensis X Soyabean pest

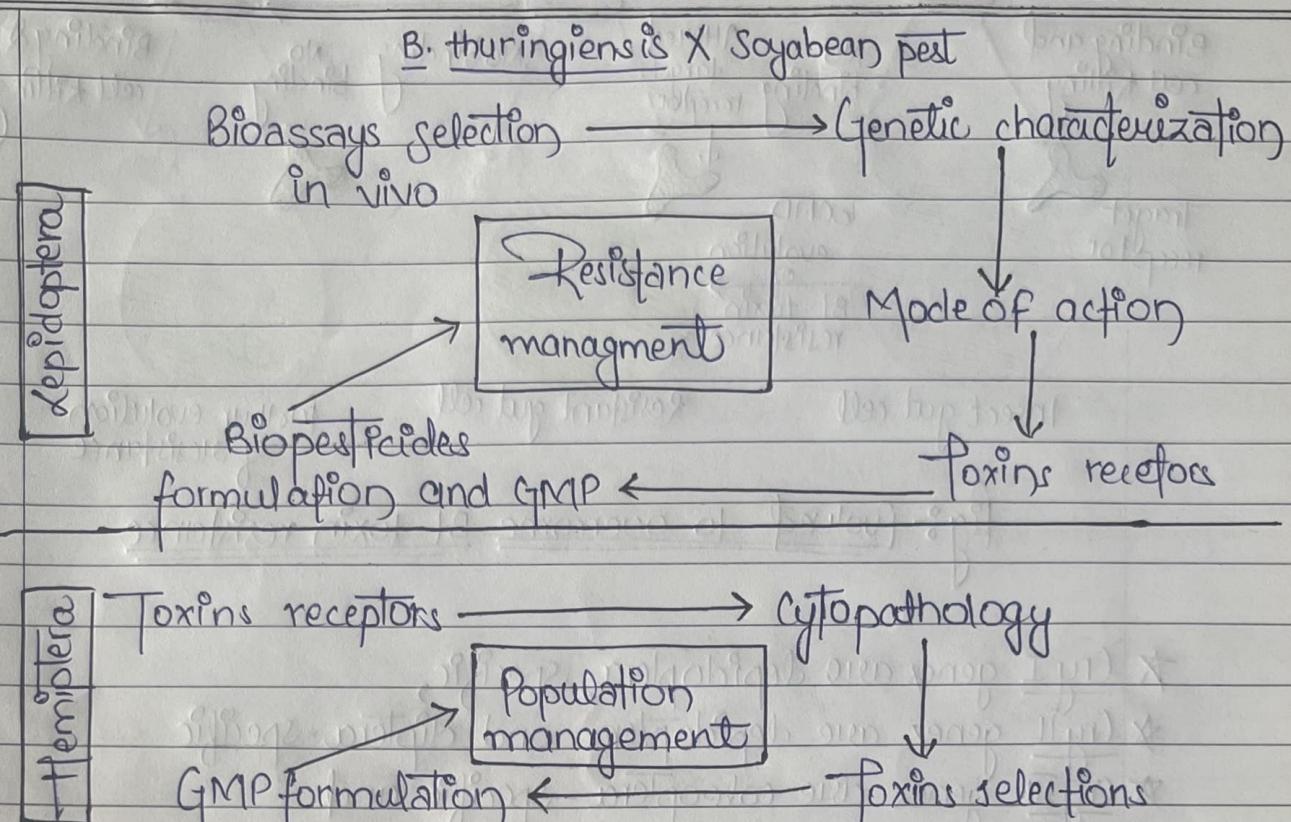
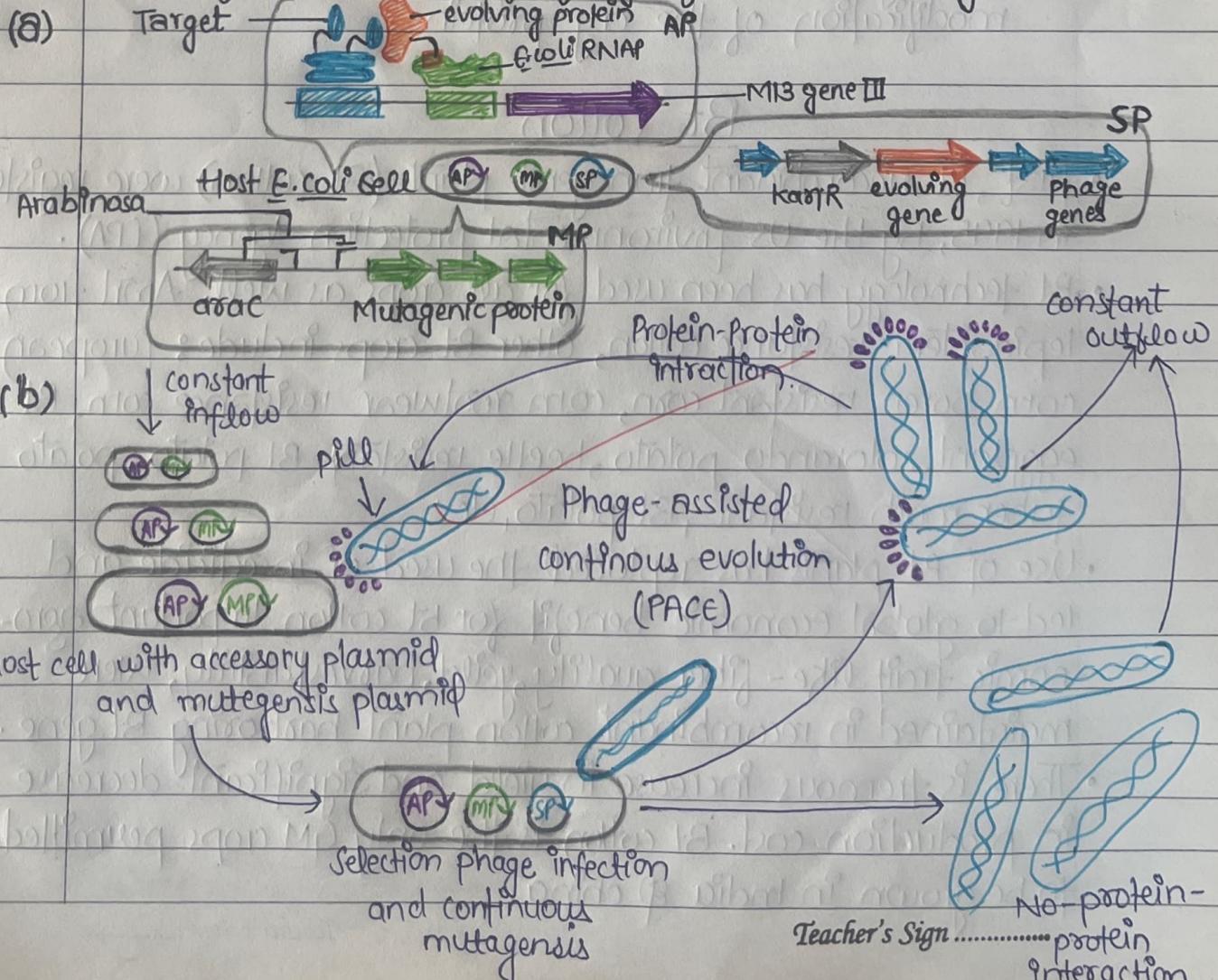
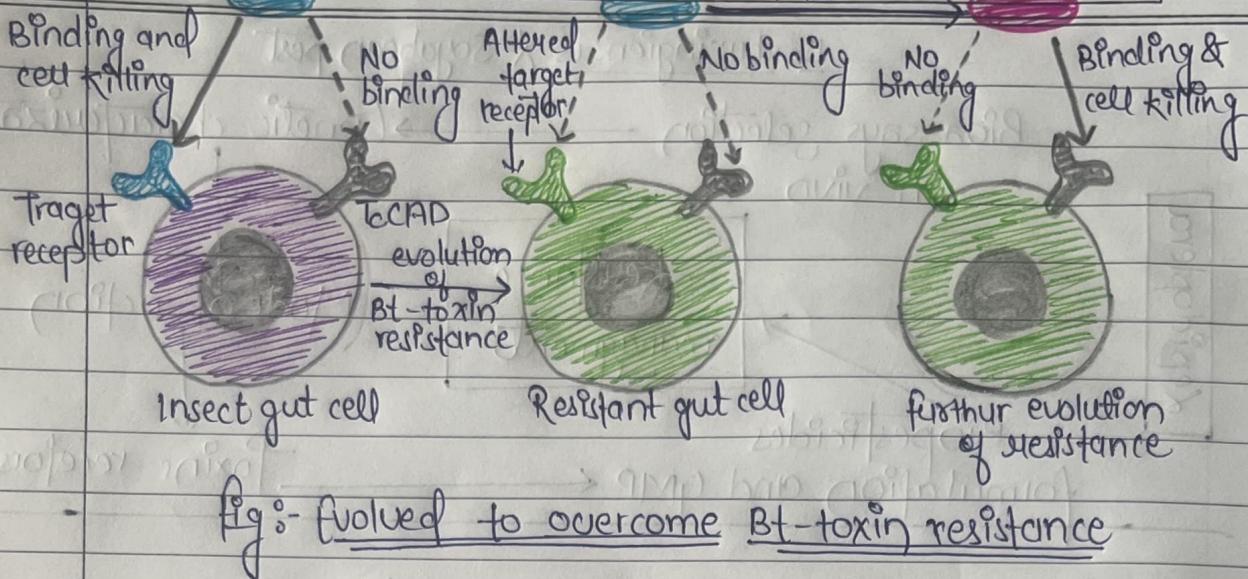


fig:-

Mode of Action and Specificity of *Bacillus thuringiensis* Toxins





- ★ Cry I genes are Lepidoptera specific,
- ★ Cry II genes are Lepidoptera and diptera-specific
- ★ Cry III genes are coleoptera-specific and
- ★ Cry IV genes are diptera-specific.

→ The toxicity of the protein changes with the molecular modification of the cry protein.

Bt Cotton

- first, Bt recombinant transgenic plant (Bt cotton) were registered in 1995, by the US Environmental protection Agency (EPA).
- Bt technology has been used for all crops as well. Apart from lepidoptera resistance cotton, other Bt crops include: european corn borer resistant corn, corn rootworm resistance corn, Bt eggplant, colorando potato, beetle resistant Bt potato, potato tuber moth resistant Bt potato, etc.
- Use of Bt plants has replaced the use of insecticide and led to global economic benefit for Bt cotton. Important agro-nomic trait like - fiber quality, yield, harvestability, were maintained in recombinant cotton plant harboring Bt gene.
- There is tremous increase in yield and significant decrease in production cost. Bt cotton is the only GM crops permitted to be grown in India & China.

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- Bt cotton is a cotton variety that has an inbuilt mechanism to protect itself from caterpillar pests, as known as the bollworm.
- The protection comes from a scientific process known as genetic modification (GM). The inbuilt mechanism is from a common soil bacterium also known as Bt, which stands for Bacillus thuringiensis.
- Bt produces a protein that is harmful for the digestion system of a caterpillar pest. When the caterpillar feeds on the Bt cotton plant, its digestive system is weakened, making it unstable to feed and it eventually dies.
- However, Bt is specific and not harmful for humans and other animals. It has been used in organic farming as a spray for over 50 years to control insect pests.

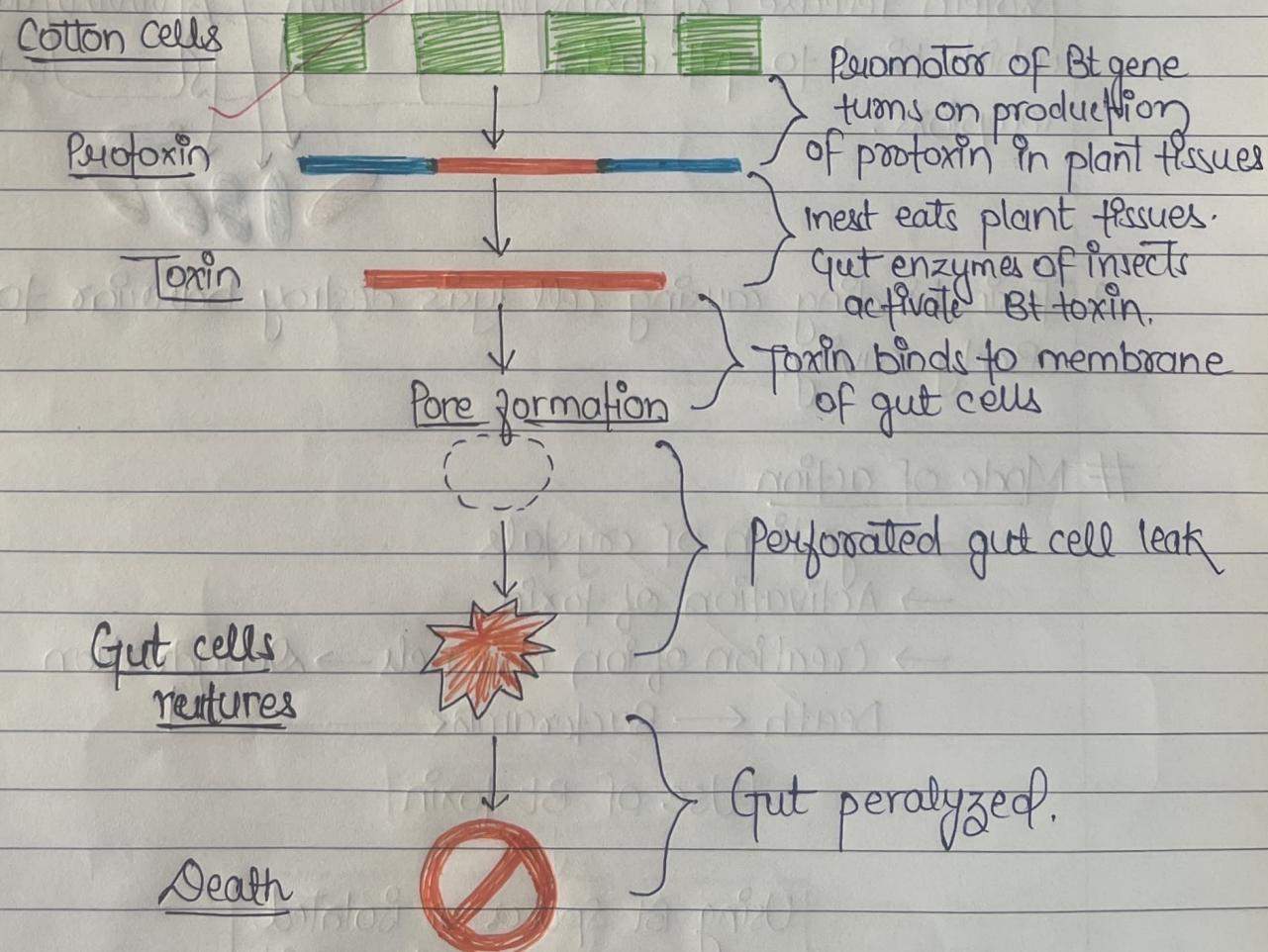


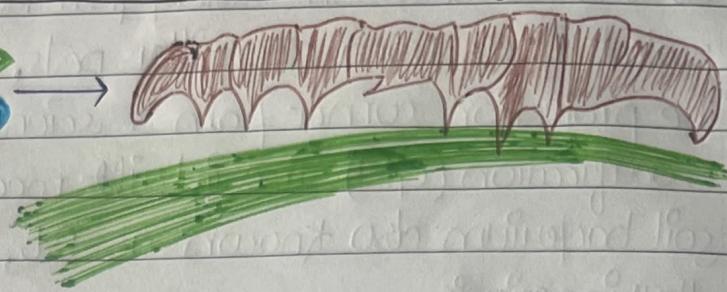
Fig: Mechanism of action of Bt Cotton

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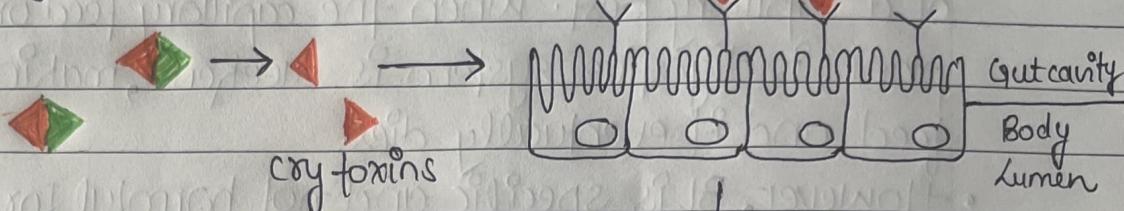
→ How does Bt cotton kill bacteria?

crystal protein

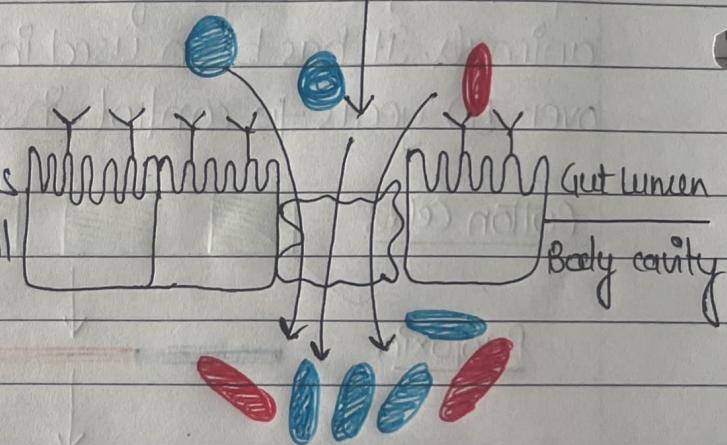
Bt spores



1. Larvae ingest Bt spore and cry protein



2. In larval midgut, proteolytic digestion of protein release cry toxins which bind to epithelial receptors.



3. Toxin binding causing cell lysis destroy barrier to Body cavity.

Mode of action

→ Ingestion of crystals

→ Activation of toxin

→ Creation of ion channels → Dehydration

Death ← Bacteremia

Use of Bt toxin

Using Bt spores or isolated crystals as biopesticide

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EDIBLE VACCINES

- Edible vaccines are vaccines produced in plant that can be administered directly through that ingestion of plant material containing the vaccine. eating the plant would then confer immunity against disease.
- One focus on current vaccines effort is on hepatitis B. Transgenic tobacco and potato were engineered to express hepatitis B. virus vaccine.
- Potato has been studied using a portion of the E. coli enterotoxin in mice and human and then transgenic potatoes were produced.
- Other candidates for edible vaccines include banana and tomato.
- In the edible vaccines, Transgenic plants are used as vaccine production systems.
- The genes encoding antigens of bacterial and viral pathogens can be expressed in plants in a form in which they retain native immunogenic properties.
- Initially thought to be useful only for preventing infectious disease, it has also found application in preventing of autoimmune disease, birth control, cancer therapy, etc.
- Edible vaccines are currently being developing for a no. of human and animal disease.
- As hippocrates said, 'Let the food be thy medicine'.

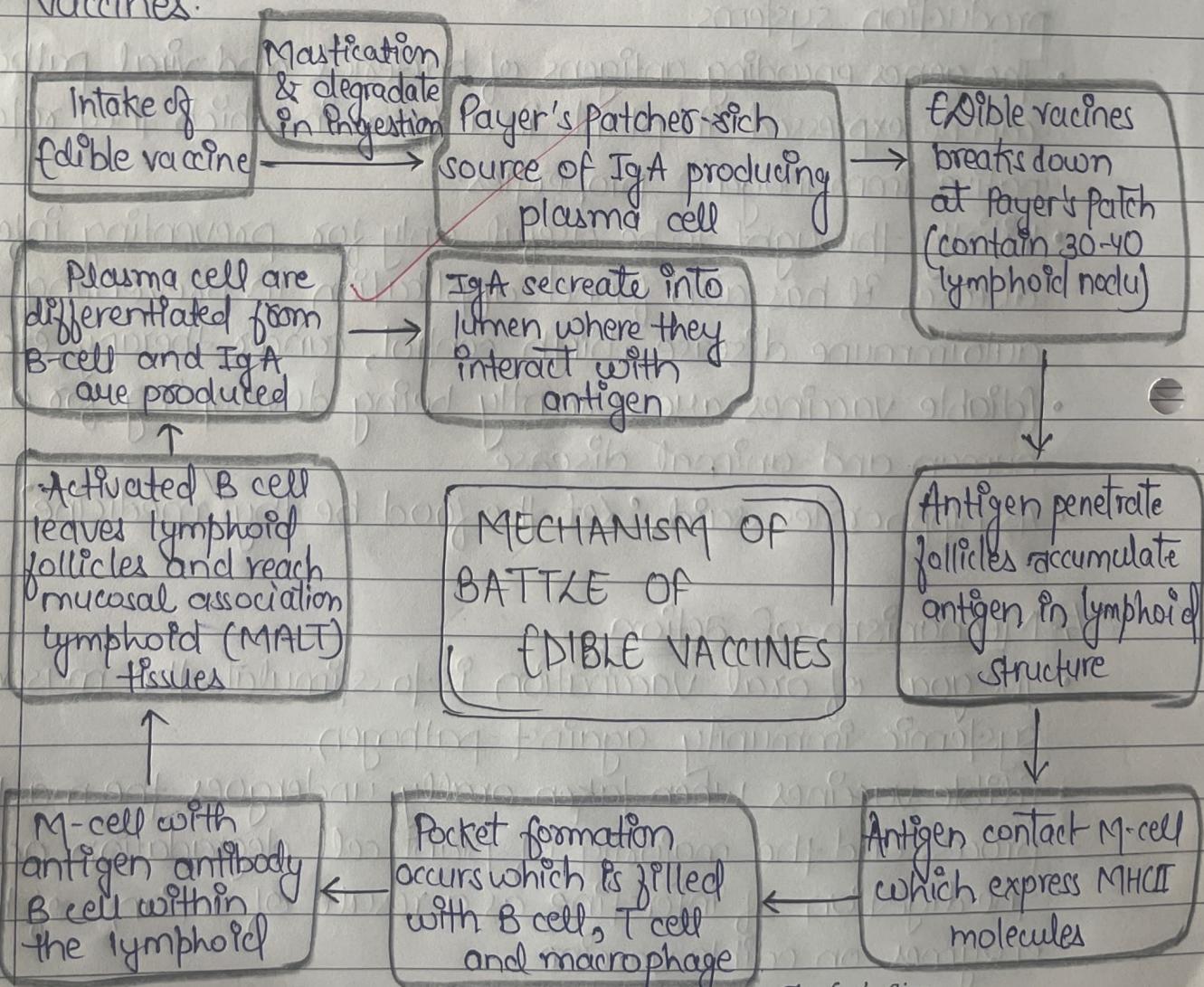
Mechanism of action

- The goal of oral vaccination is to stimulate the mucosal and systemic immunity against pathogen.
- Edible vaccines when taken orally undergoes the mastication process and the majority of plant cell degradation occur in the digestion of intestine as a result of action of bacterial enzymes on edible vaccine.

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- Peyer's patches (pp) are an enriched source of IgA producing plasma cell and have the potential to populate mucosal tissues and serve as mucosal immune effector site.
- The breakdown of edible vaccine near PP, consisting of the 30-40 lymphoid nodules on the outer surface of intestine and contain follicles.
- These follicles act as the site from which antigen penetrate the intestinal epithelium, thereby antigen accumulation antigen within organized lymphoid structure.
- Then antigen comes in contact with M-cell
- M cell passes the antigen to macrophages & B cell
- These B cell activates the T cell to produce immune response
- In this way, the immunity is activated by the edible vaccines.



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Developing an edible vaccines.

- There are Two ways of developing edible vaccines:-
- In one case, the entire structural gene is inserted in plant transformation vector between 5' and 3' regulatory element, the will allow the transcription and accumulation of encode sequence in the plant
- In second case, entipode within the antigen are identity, DNA fragment encoding these can be used to construct gene by fusion with a protein coat gene from plant virus. e.g. TMV.

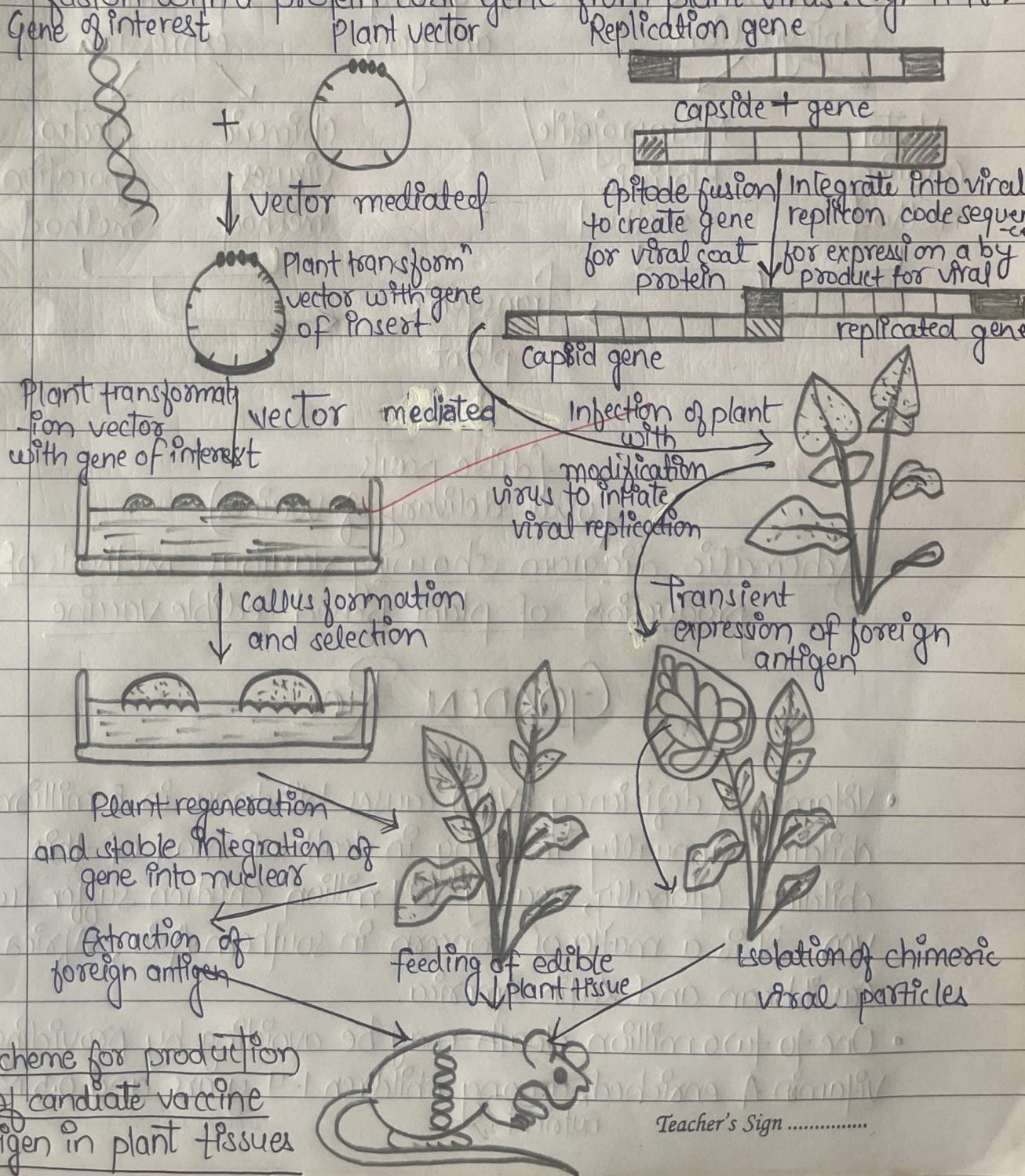
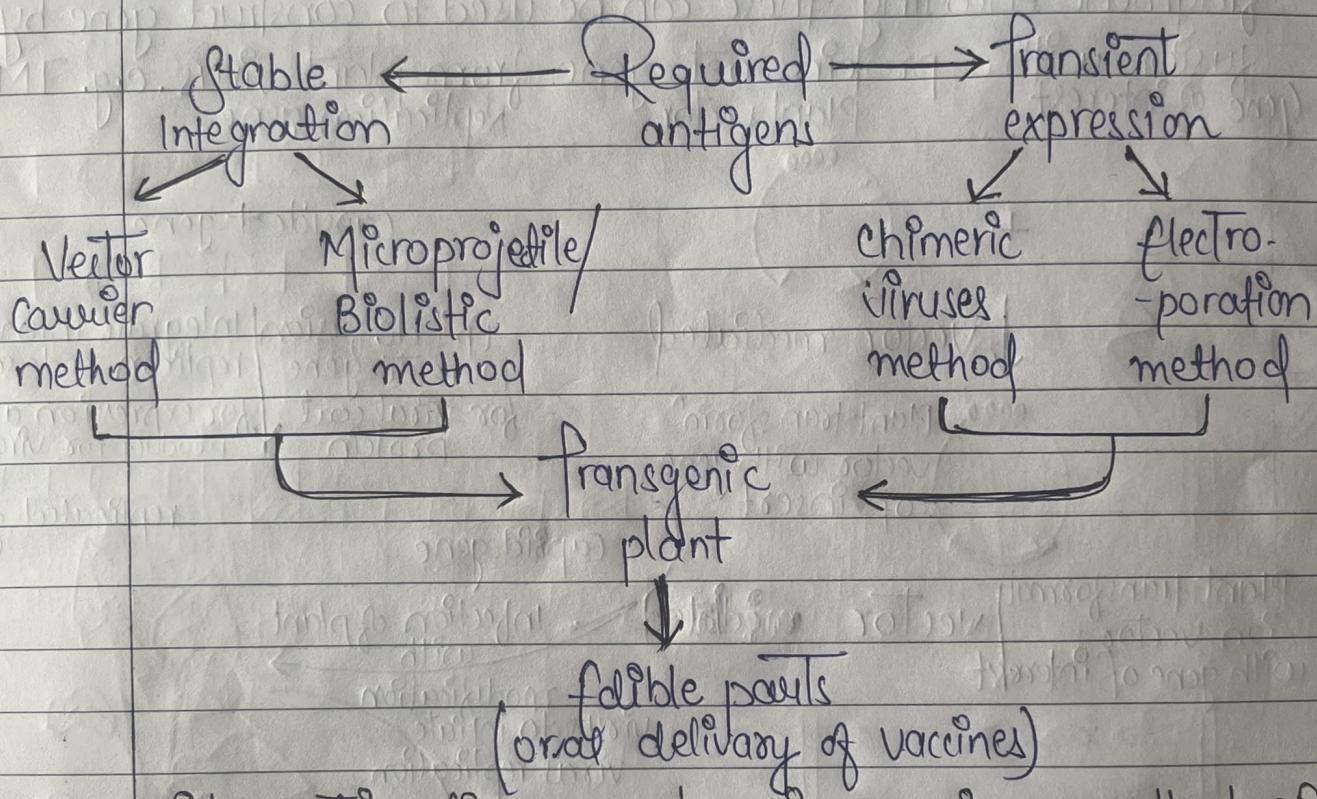


fig: Scheme for production
of candidate vaccine
antigen in plant tissues

Teacher's Sign

IDEAL VACCINES

- It should not be toxin or pathogenic
- Low level of side effects
- It should not contaminated the environment
- It should not cause problem in individual
- Technique of vaccine should be simple
- It should be cheap.



Schematic diagram showing various methods of production of plant based edible vaccine

GOLDEN RICE

- Vitamin A deficiency (VAD) causes blindness in 5 million children annually resulting in death of most of these children. Globally, nearly 124 million children suffer from VAD. It is a matter of concern in southern Asia, Africa, Caribbean and Latin America.
- One to two million deaths may be avoided by providing Vitamin A enriched among children 1-4 years and 0.5 M.

- during high age group of children. One approach providing Vitamin A capsules to children and new mothers and alternative provitamin A can be provided in the form of b-carotene in rice which is one of the best known examples of nutritional improvement of a food crop.
 - Carotenoids are compound belonging to a class of plant metabolise called terpenoids or isoprenoids. Carotenoid are produced from it precursor by a biochemical pathways located in plasmid.
 - The 40-carbon backbone of b-carotene (pro vitamin-A) is known as phytoene and it assemble by two 20-carbon geranylgeranyl diphosphate (GGPP) molecules by the enzyme phytoene synthase.
 - Double bond in phytoene are added by denaturation steps producing an antioxidant compound called lycopene, give red colour to tomatoes which is later converted to b-carotene by the enzymes lycopene cyclase.
 - Within the aim of fortified food production, research was carried out to grow & consume variety of rice in areas where VAD is found to be common.
 - A variety of Oryza sativa was produced through genetic engineering capable of synthesizing beta carotene, a precursor of vitamin A in the endosperm of rice (Transgenic). The variety known as 'Golden rice', differ from the parental variety by having two additional beta-carotene synthesis genes.
- ★ Golden rice produced by transforming rice with two beta-carotene synthesis genes:

- (1) psy (phytoene synthase) from Baifodii
- (2) crt I (carotene desaturase) from a soil bacterium.

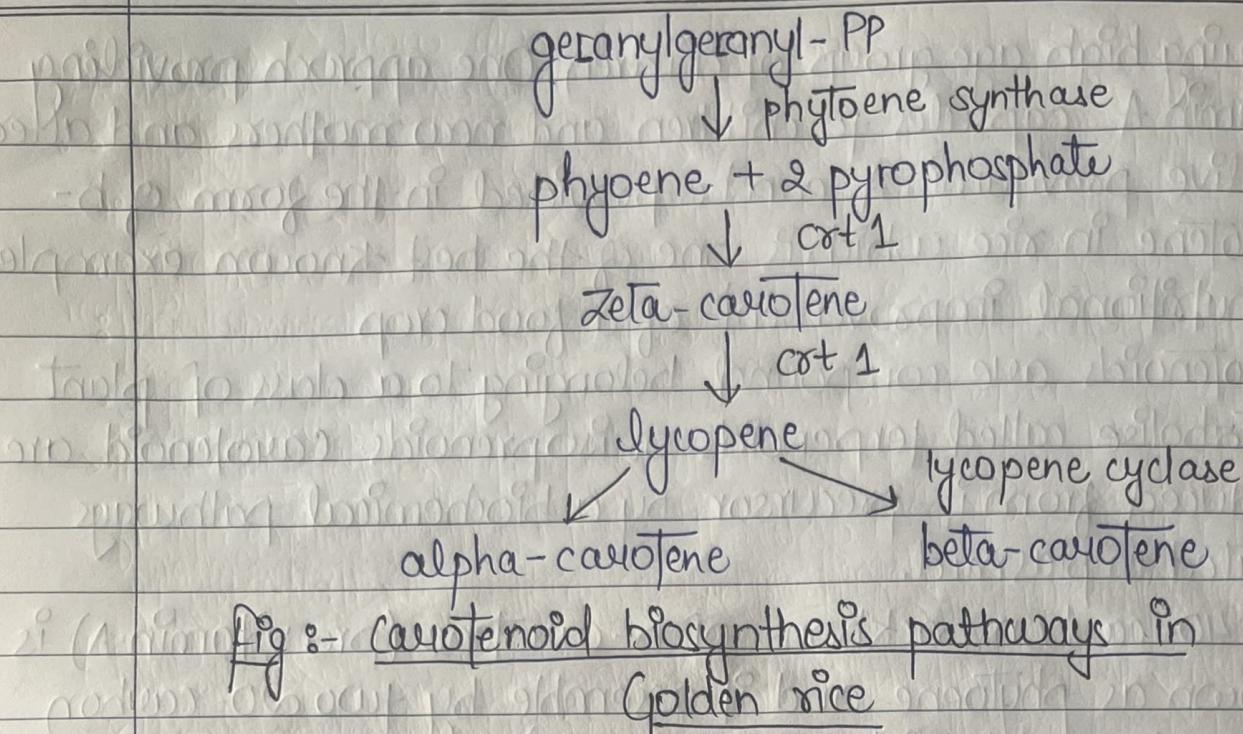


Fig :- carotenoid biosynthesis pathways in Golden rice

- Both these genes are inserted into rice nuclear genome under the control of an endosperm specific promoter to express them only in endosperm. The end product of this pathway is lycopene, but plant don't accumulate it otherwise the rice would have been red.
- An endogenous enzymes inside the rice convert lycopene to beta-carotene giving yellow colour after which it is named. The detail of golden rice is first published in 2000. This is product of an eight year project by Ingo Potrykus of Swiss Federal Institute of Technology and Peter Beyer of University of Freiburg.
- In the year 2005, a new variety of rice, called golden rice 2, producing 23 times more beta-carotene in comparison of original golden rice, was announced.

Topic.....

Date.....

Rice endosperm

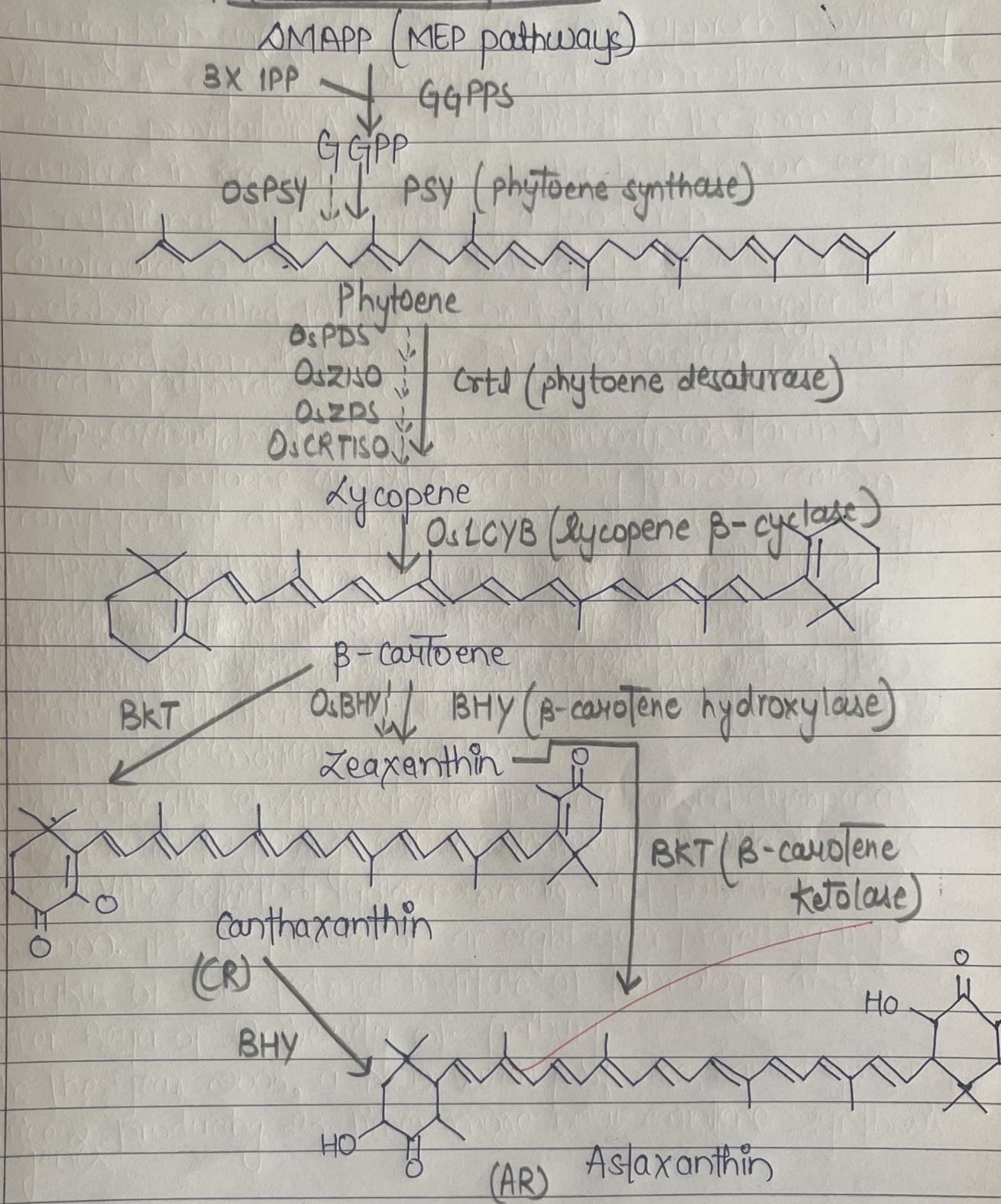


Fig :- flow diagram for chemical pathways in golden rice

No marks

Teacher's Sign